



The IWBBIO 2022 (9th International Work-Conference on Bioinformatics
and Biomedical Engineering)

Integrative Analysis of Ovarian Serious Adenocarcinoma to Understand Disease Network Biology



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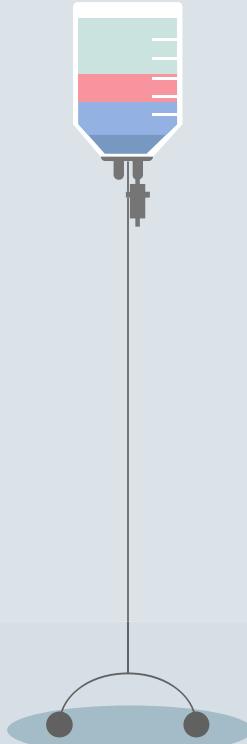
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“He who conceals his disease cannot expect to be cured !”

—An Ethopian Proverb



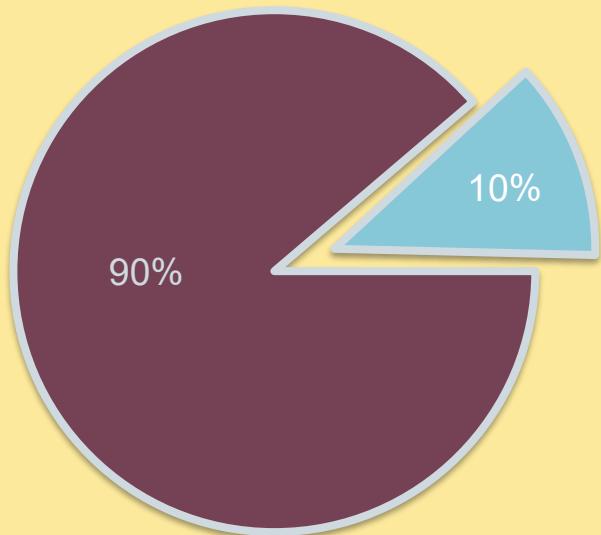
INTRODUCTION

Ovarian Cancer (OC)

- Ovarian cancer (OC) is one of the gynecologic oncologic malignancies in females that leads to lethality annually is a cancer that develops on the surface of the ovary.
- As per the NCRP-ICMR (2013) it is known to be the third leading cause of cancer triggered deaths in females and henceforth is tagged as a ***Silent Killer***.
- Ovarian Cancer has majorly five subtypes, namely - ***High grade serous*** (HGS-OvCa), ***Low grade serous*** (LGS- OvCa), ***Endometrioid***, ***Mucinous***, ***Clear Cell Tumors***.
- High grade serous is the most common types of ovarian cancer (OC) in females in India.



TYPES OF OVARIAN CANCER



90%

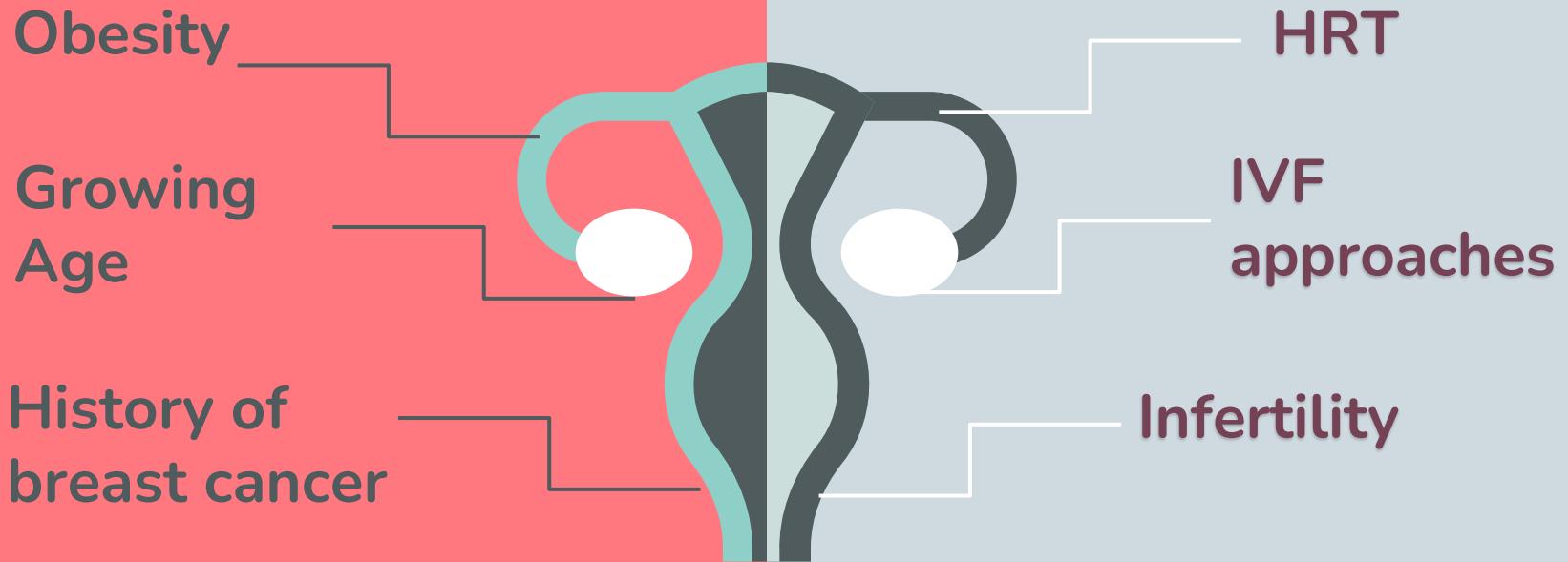
Are epithelial carcinoma



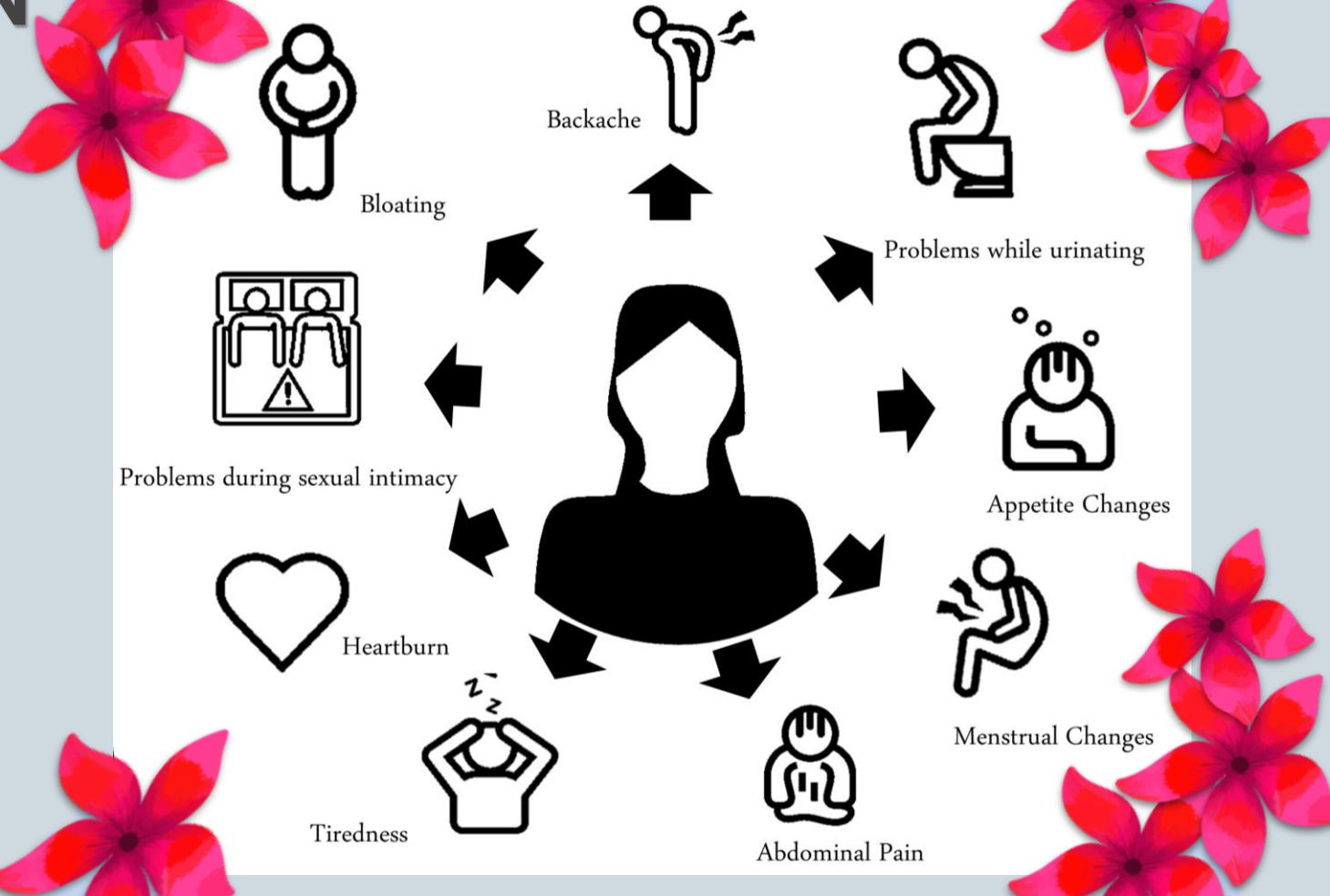
10%

Are germ cell and stromal tumors

FACTORS INVOLVED IN OVARIAN CANCER

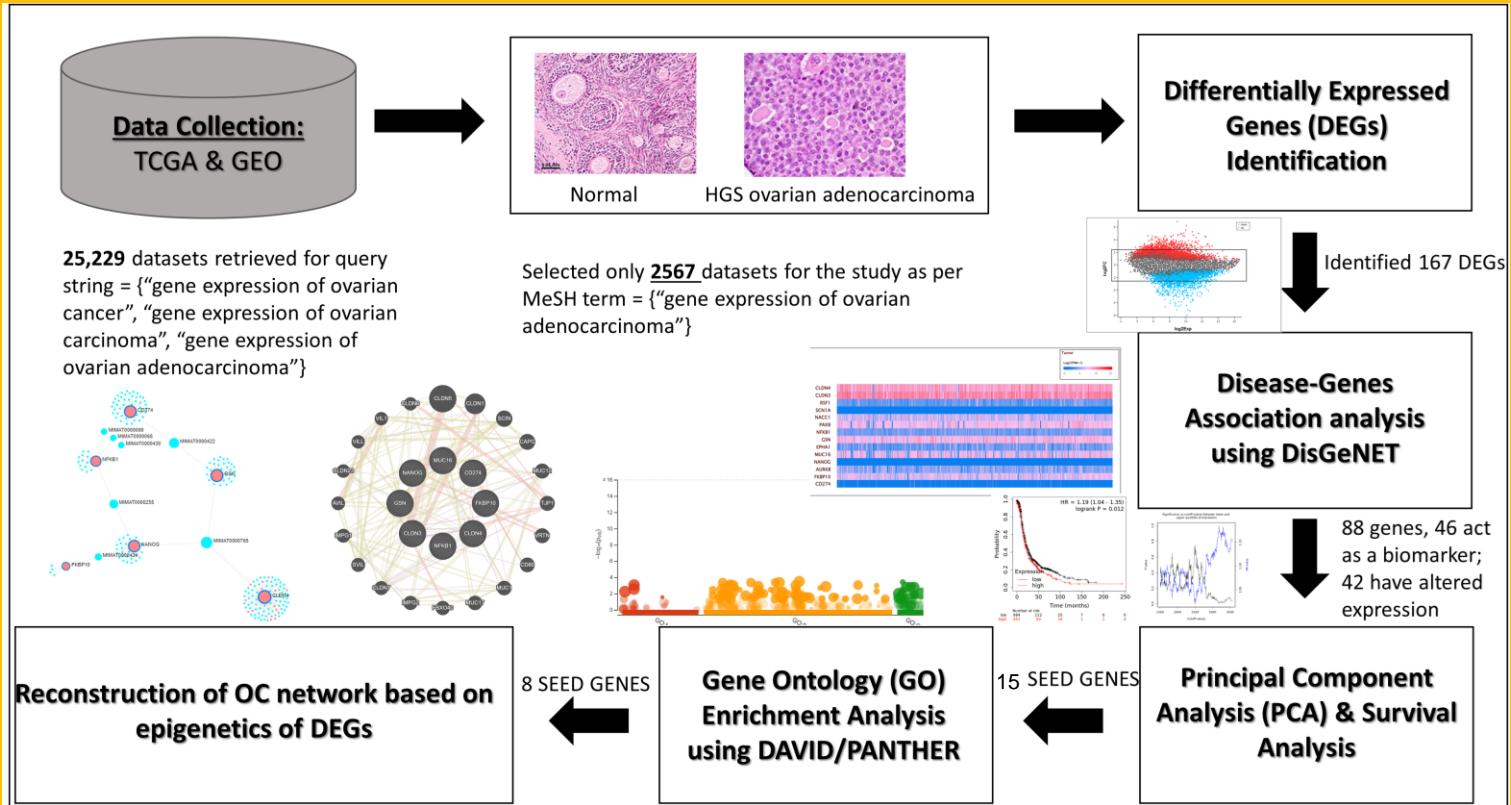


COMMON SIGNS



OBJECTIVE

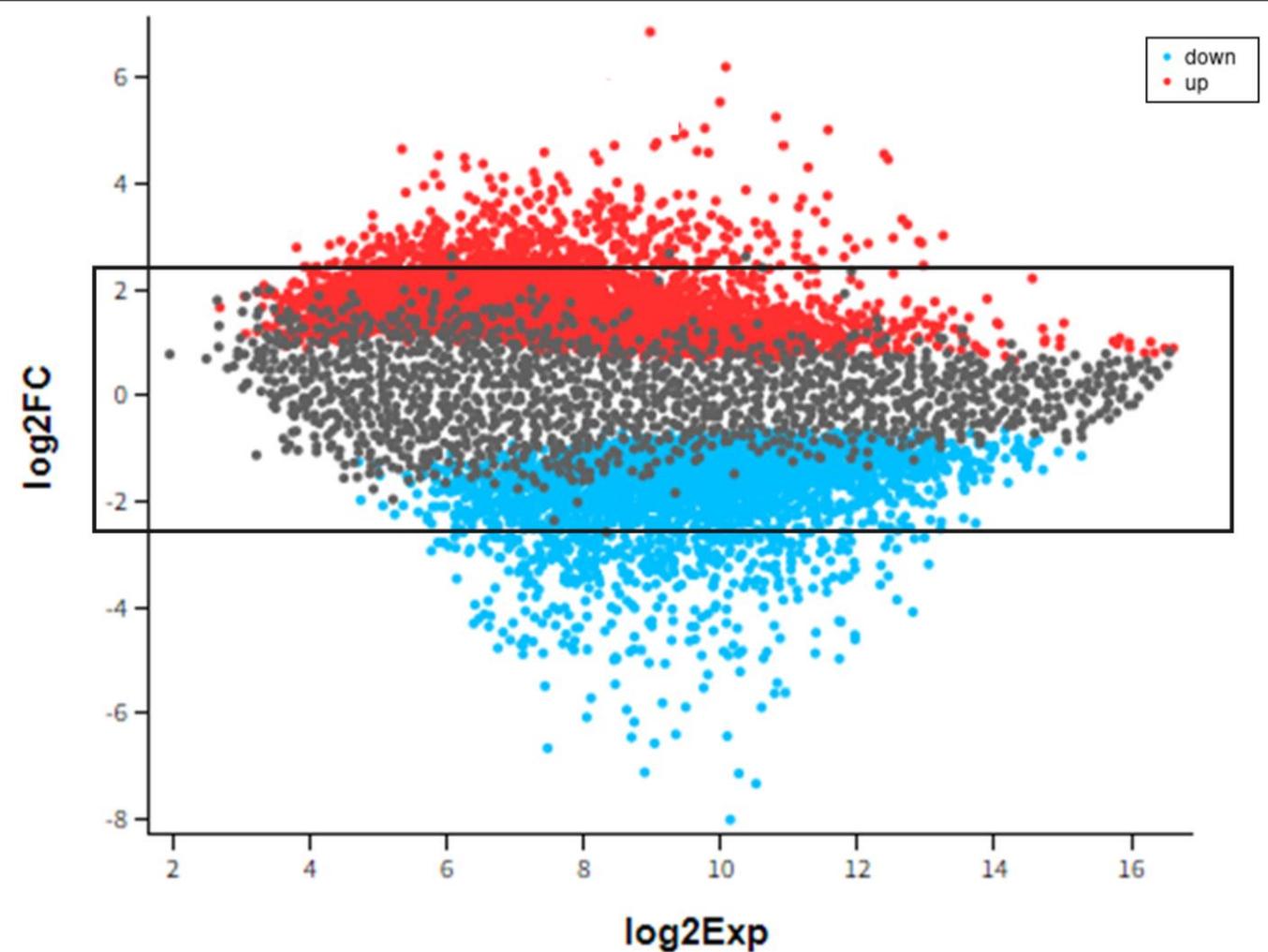
**Analysing the
behaviour of ovarian
cancer (OC) network**



DATABASES/SOFTWARE/PLUGINS USED

S.No.	NAME	PURPOSE	LINK
1.	The Cancer Genome Atlas (TCGA) database	OVCA gene expression dataset retrieval	https://www.cancer.gov/about-nci/organization/ccg/research/structural-genomics/tcga
2.	Gene expression omnibus (GEO) database	OVCA gene expression dataset retrieval	https://www.ncbi.nlm.nih.gov/geo/
3.	R Studio (packages used: GeoTcgadata, BiocManager,forcats, stringr,igraph ggplot2, ggrepel, readr, tidy, survminer, GEOquery, limma, pheatmap, ClustVis, pcaExplorer, OmicsNetR)	Differentially expressed genes (DEGS) identification, inspection of clinical variables, sample clustering and principal component analysis (PCA), network visualization	https://www.r-project.org/
4.	Kaplan Meier (KM) survival estimation webserver	Prediction of survival estimates of the seed genes.	https://kmplot.com/analysis/index.php?p=background
5.	Cytoscape software (plugins used: DisGeNET, BINGO, GeneMania & EnrichmentMap)	Network visualization, disease-gene associations, gene ontology (GO) representation in network, gene regulatory network re-construction and pathway enrichment	https://cytoscape.org/

SCATTER PLOT



Here, the red-colored dots represent **up-regulated** (over-expressed) genes while dots in blue represent **down-regulated** (under-expressed) genes in serous ovarian adenocarcinoma.

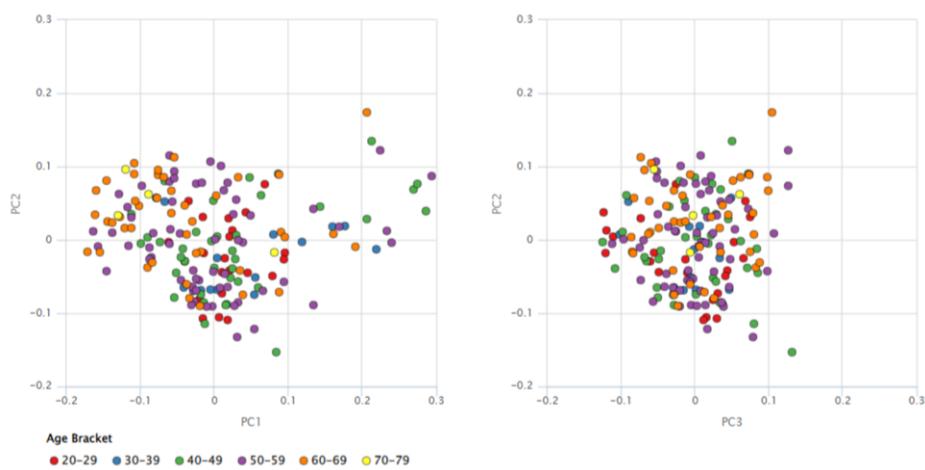
The grey dots represent those genes that maintain a normal level of expression.



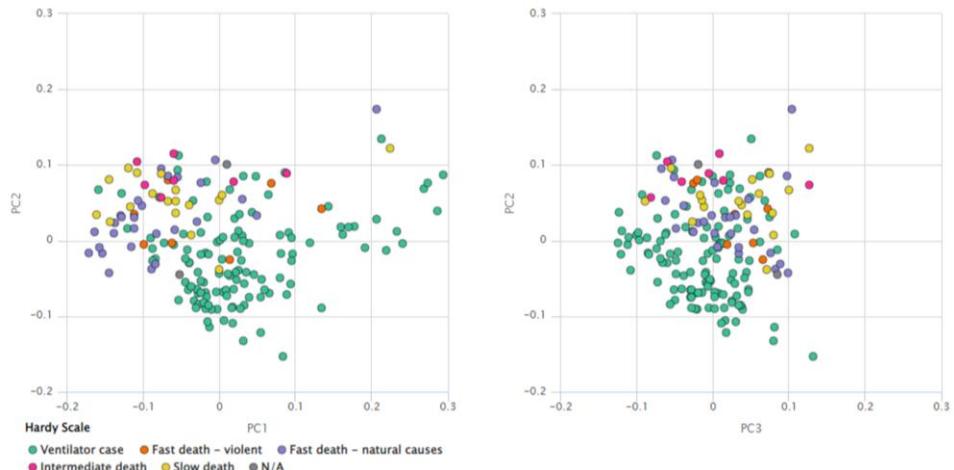
DISEASE-GENE ASSOCIATION USING DiSGeNET

S.No.	Seed Gene	Association	Score
1.	<i>CLDN3</i>	Biomarker	0.02
2.	<i>CLDN4</i>	Biomarker	0.11
3.	<i>RSF1</i>	Altered Expression	0.02
4.	<i>SCN1A</i>	Biomarker; Altered Expression	0.3
5.	<i>NACC1</i>	Biomarker, Altered Expression	0.01
6.	<i>PAX8</i>	Biomarker	0.1
7.	<i>NFKB1</i>	Biomarker	0.01
8.	<i>GSN</i>	Biomarker, Altered Expression	0.1
9.	<i>EPHA1</i>	Biomarker	0.02
10.	<i>MUC16</i>	Biomarker	0.3
11.	<i>TP53</i>	Biomarker	0.1
12.	<i>NANOG</i>	Altered Expression	0.01
13.	<i>AURKB</i>	Altered Expression	0.3
14.	<i>FKBP10</i>	Altered Expression	0.01
15.	<i>CD274</i>	Altered Expression	0.02

PRINCIPAL COMPONENT ANALYSIS (PCA)

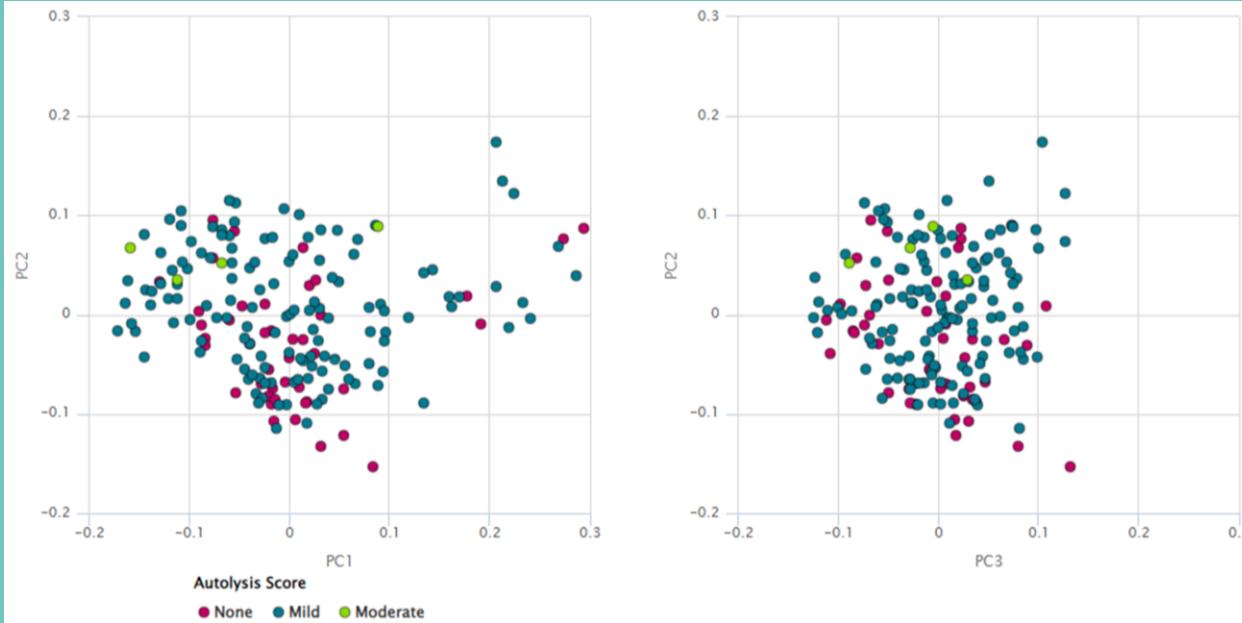


a) PCA representing an incidence of ovarian serous adenocarcinoma based on the age bracket of a female.



b) PCA representing the severity of patients based on Hardy-Weinberg's equilibrium.

PRINCIPAL COMPONENT ANALYSIS (PCA)

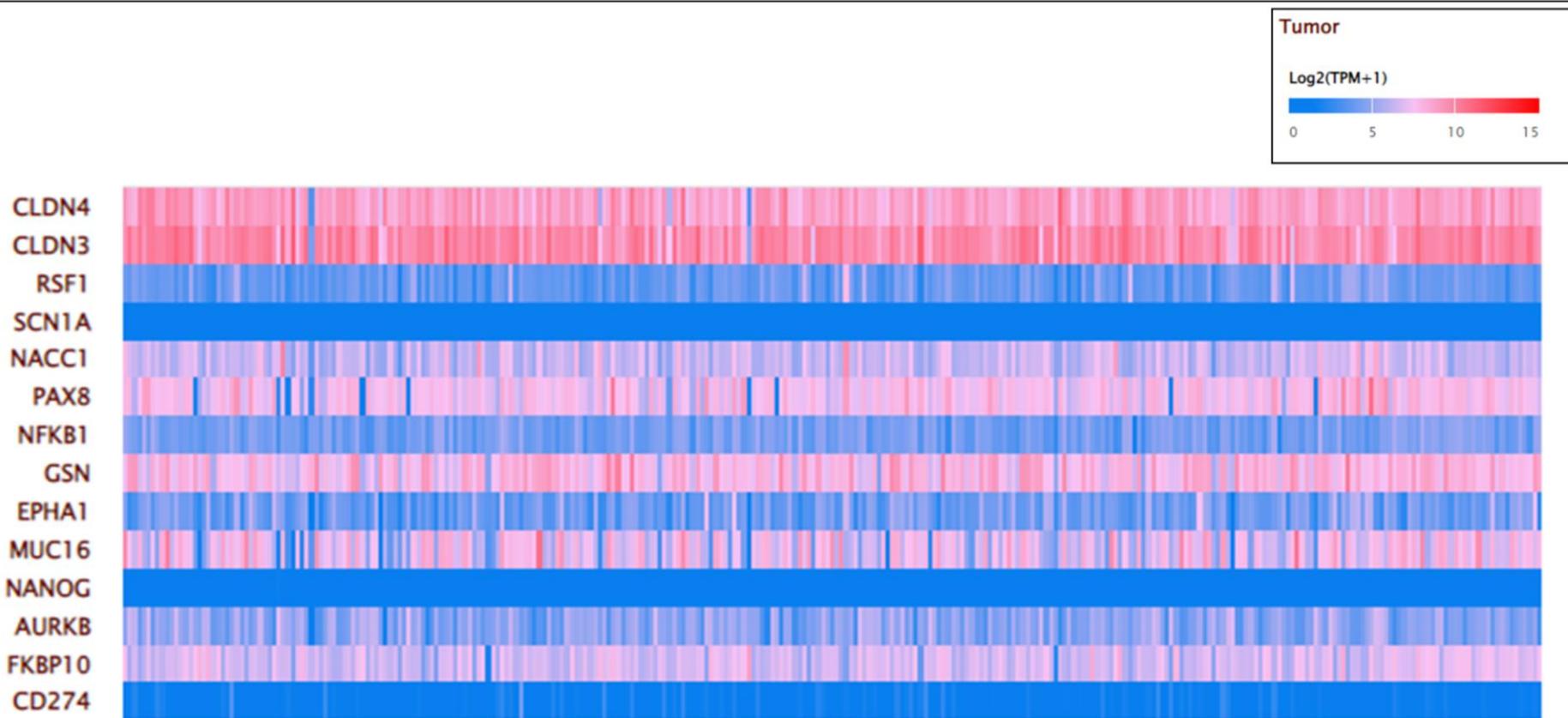


c) PCA representing autolysis of genes involved in ovarian serous adenocarcinoma.

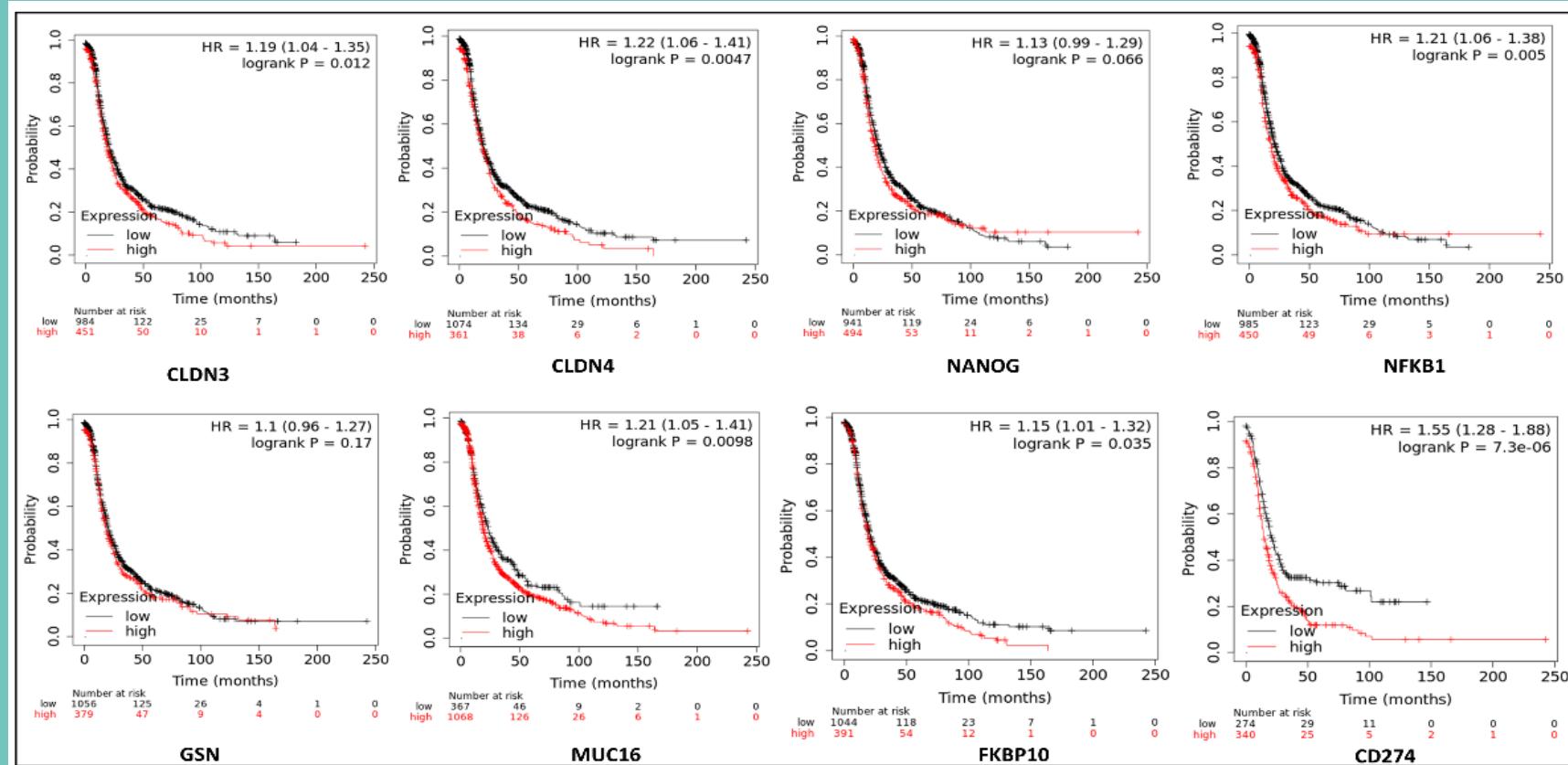
MEDIAN SURVIVAL ESTIMATES FOR 15 SEED GENES

S.No.	Seed Gene	Description	Low expression cohort (months)	High expression cohort (months)
1.	<i>CLDN3</i>	Claudin 3	20.63	18.83
2.	<i>CLDN4</i>	Claudin 4	20.2	19.0
3.	<i>RSF1</i>	Remodeling and spacing factor 1	18.0	15.0
4.	<i>SCN1A</i>	Sodium voltage-gated channel alpha subunit 1	18.0	21.29
5.	<i>NACC1</i>	Nucleus Accumbens Associated 1	18.27	16.0
6.	<i>PAX8</i>	Paired box gene 8	18.79	22.5
7.	<i>NFKB1</i>	Nuclear factor kappa B subunit 1	21.13	17.9
8.	<i>GSN</i>	Gelsolin	20.53	18.43
9.	<i>EPHA1</i>	Ephyrin type A receptor 1	18.23	22.02
10.	<i>MUC16</i>	Mucin 16	23.24	19.0
11.	<i>TP53</i>	Tumor protein P53	17.43	21.29
12.	<i>NANOG</i>	Nanog Homeobox	21.13	18.2
13.	<i>AURKB</i>	Aurora Kinase B	18.93	20.63
14.	<i>FKBP10</i>	FKBP Prolyl Isomerase 10	20.47	19.0
15.	<i>CD274</i>	CD274 molecule	20.0	14.37

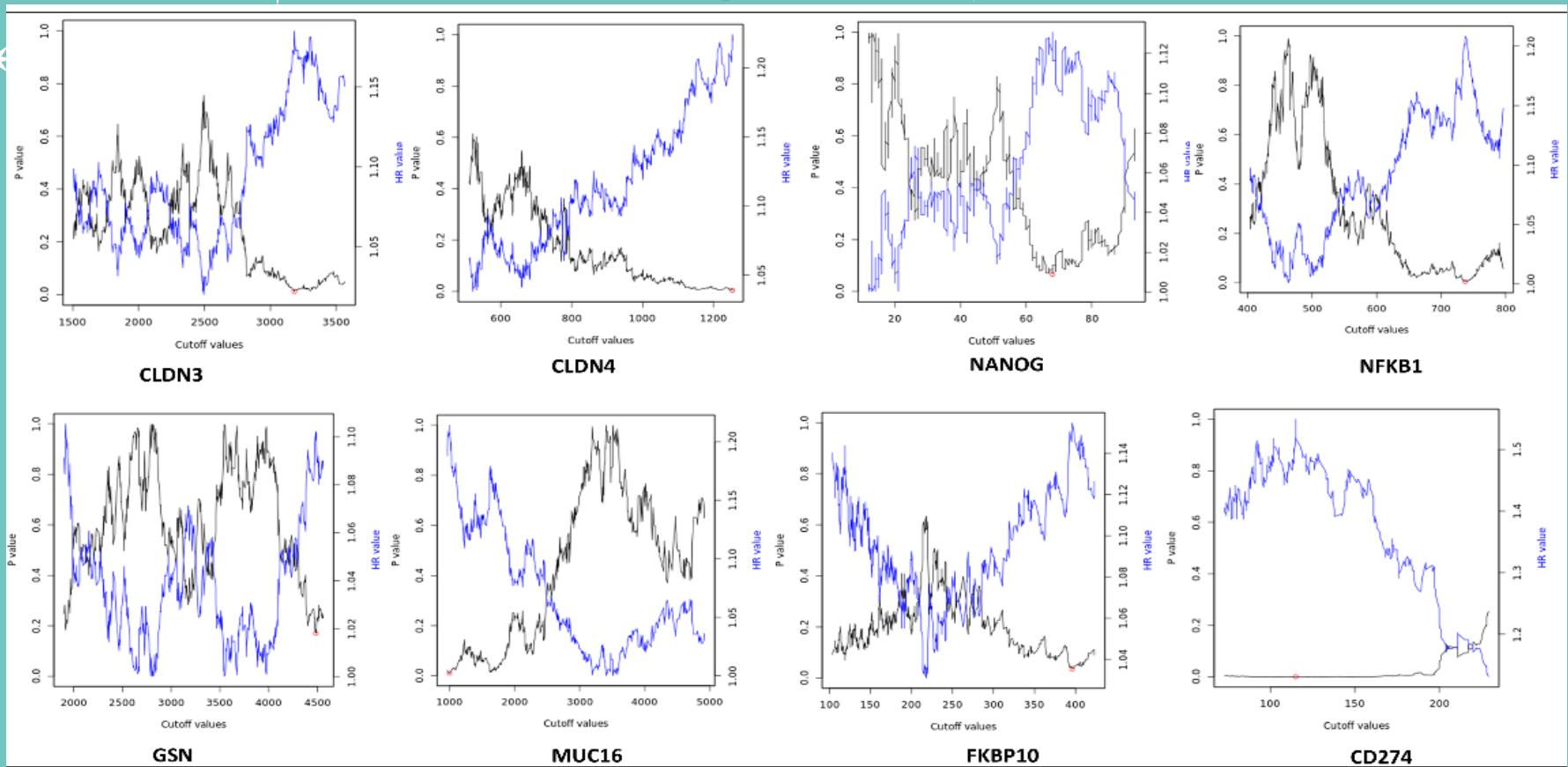
EXPRESSION PATTERNS OF SEED GENES IN THE FORM OF HEATMAP



KAPLAN MEIER SURVIVAL CURVES FOR TOP 8 GENES



KM Significance vs Cut-Off Values between Upper & Lower Quartiles of Expression



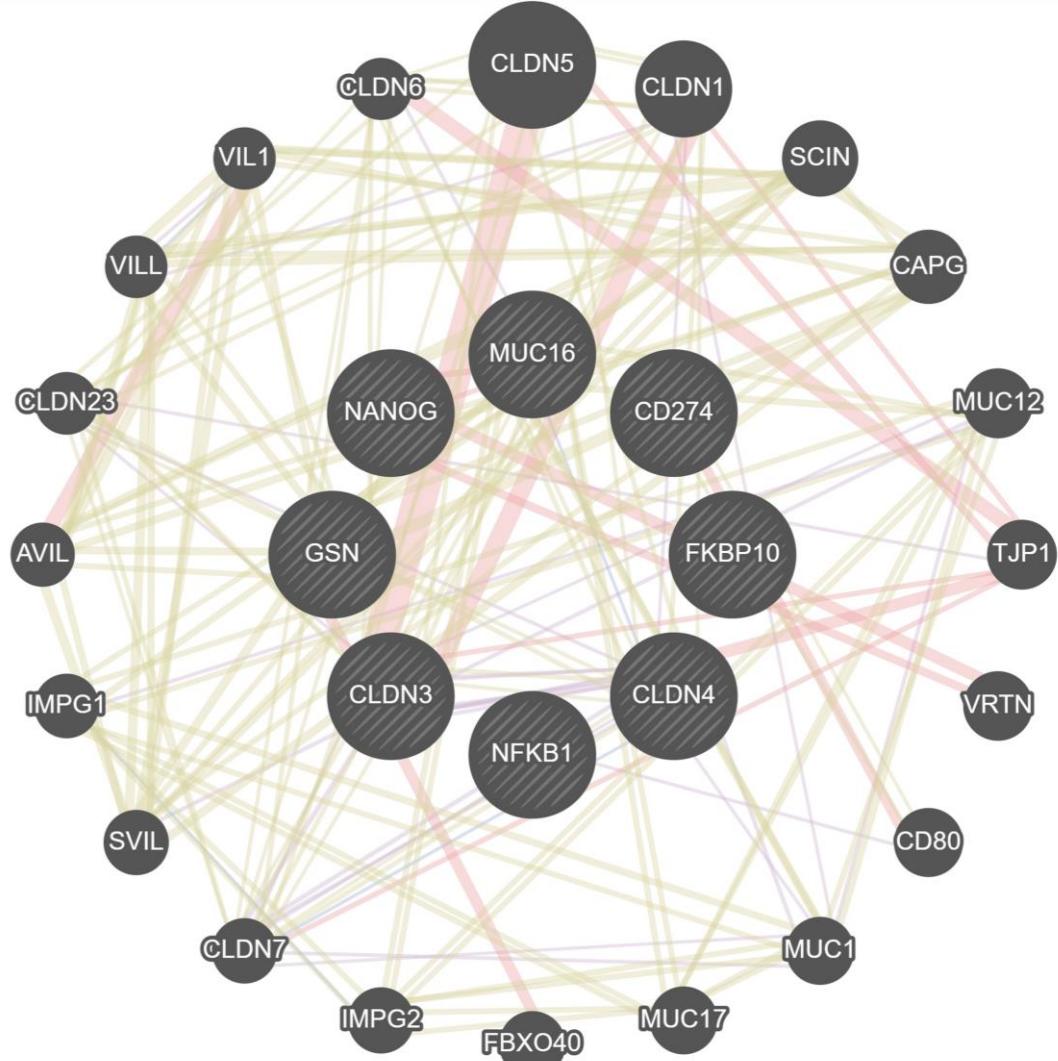
★ GENE ONTOLOGY ENRICHMENT OF TOP 8 GENES

GENE ONTOLOGY (GO)				
Seed Gene	Biological Processes	Molecular Functions	Cellular Localization	Pathway Collection
<i>CLDN3</i>	Response to stimulus,	Binding, Protein Binding,	Membrane enclosed	Proteins with altered
<i>CLDN4</i>	Wound healing, Cell adhesion,	DrugBinding, Chloride	lumen, Apicolateral	expression in endometrial
<i>NFKB1</i>	Macromolecular metabolic	channel activity, cis-trans	plasma membrane,	cancer, proteins involved
<i>GSN</i>	process, biological adhesion,	isomerase activity, DNA	Organelle lumen, Lateral	in endometriosis,
<i>MUC16</i>	Positive regulation of biological	binding transcription	plasma membrane, Tight	Coagulation, Apoptosis,
<i>NANOG</i>	process, Positive regulation of	repressor activity, Actinin	junction, Anchoring	Inflammatory response,
<i>FKBP10</i>	metabolic process, response to	binding	junction, Endomembrane	TNF-Alpha signalling via
<i>CD274</i>	cytokine, regulation of		system, Bicellular tight	NF-kB, Cancer
	peptidase activity, Tight		junction, Cellular	immunotherapy by PD-1
	junction assembly, Positive		component	blockade WP4585,
	regulation of cell motility			Interactions between
				immune cells and
				microRNAs in tumor
				microenvironment
				WP4559

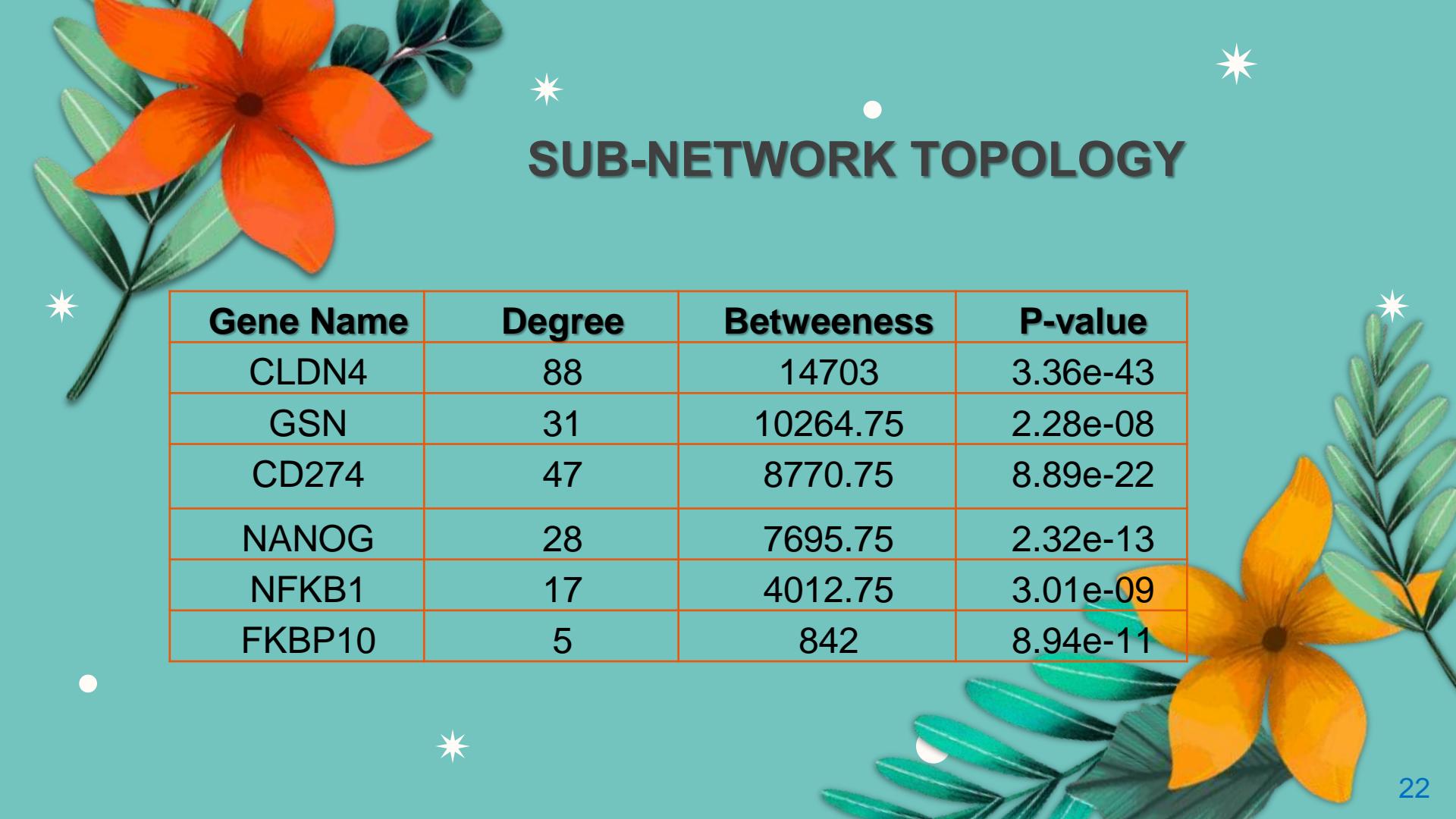




GENE REGULATORY NETWORK(GRN) CONSTRUCTION



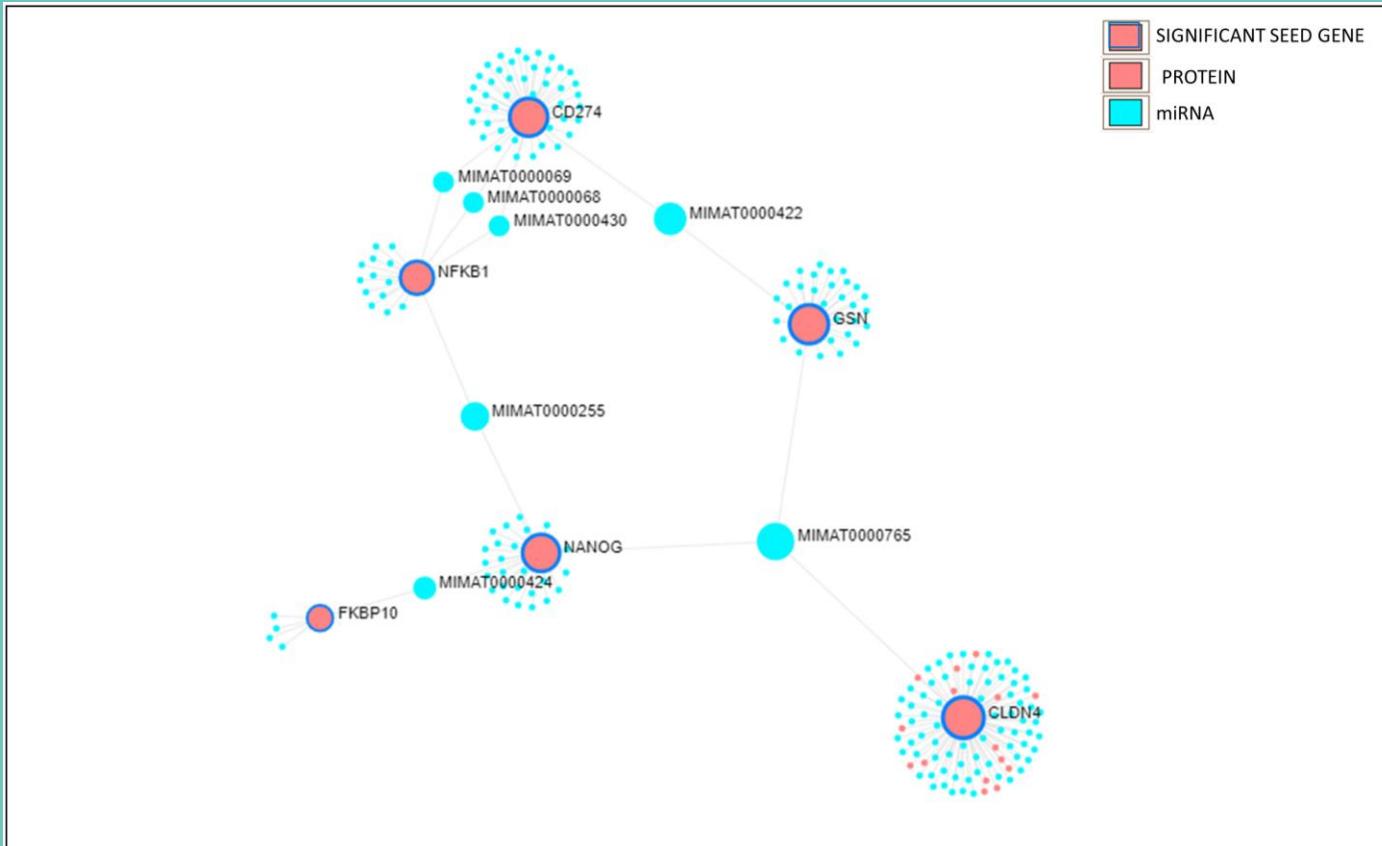
To simplify the network, we used ***label propagation algorithm*** (LPA) to visualize the GRN.



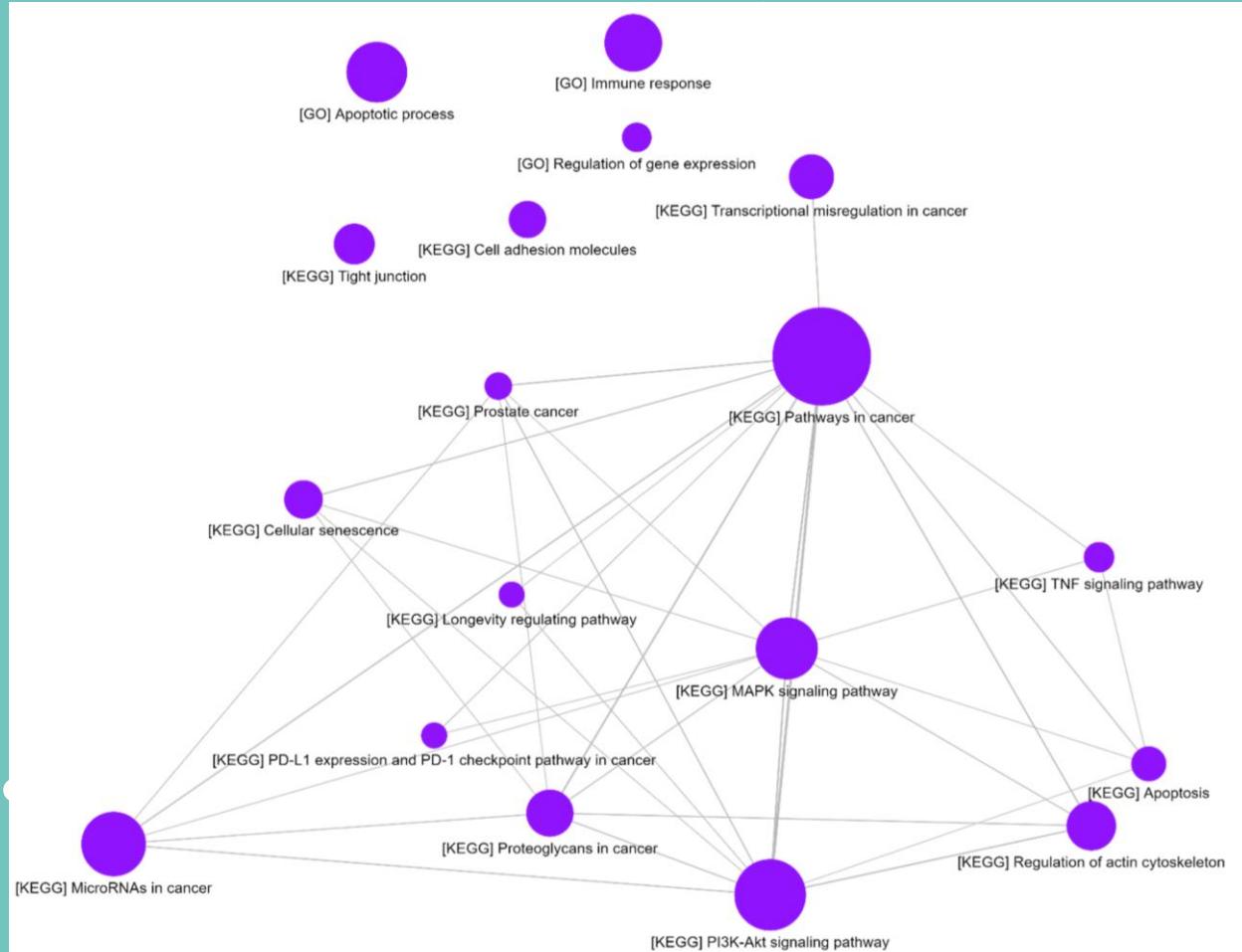
SUB-NETWORK TOPOLOGY

Gene Name	Degree	Betweeness	P-value
CLDN4	88	14703	3.36e-43
GSN	31	10264.75	2.28e-08
CD274	47	8770.75	8.89e-22
NANOG	28	7695.75	2.32e-13
NFKB1	17	4012.75	3.01e-09
FKBP10	5	842	8.94e-11

SUBNETWORK OF THE 6 HUB GENES – CD274, NFKB1, NANOG, FKBP10, GSN, CLDN4.



SIGNIFICANT PATHWAYS WHERE THE 6 HUB GENES ARE INVOLVED



SUMMARY



15 seed genes
mainly target females of
age bracket 50–69.

CLDN4, GSN,
CD274, NANOG, FKBP10
and NFKB1 showed a
strong sub-network
associating with proteins
and microRNAs.



CLDN3, CLDN4, NFKB1,
GSN, MUC16, NANOG,
FKBP10 and CD274 are
influential in dominating
ovarian serous
adenocarcinoma.

Contributors



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REFERENCES

- JEONG, Y. J., OH, H. K., & CHOI, H. R. (2019). METHYLATION OF THE RELA GENE IS ASSOCIATED WITH EXPRESSION OF NF- κ B1 IN RESPONSE TO TNF- α IN BREAST CANCER. *MOLECULES*, 24(15), 2834. DOI:10.3390/MOLECULES24152834
- HOUSHDARAN, S., NEZHAT, C. R., VO, K. C., ZELENKO, Z., IRWIN, J. C., & GIUDICE, L. C. (2016). ABERRANT ENDOMETRIAL DNA METHYLOME AND ASSOCIATED GENE EXPRESSION IN WOMEN WITH ENDOMETRIOSIS. *BIOLOGY OF REPRODUCTION*, 95(5), 93. [HTTPS://DOI.ORG/10.1095/BIOLREPROD.116.140434](https://doi.org/10.1095/biolreprod.116.140434)
- GAN, L., YANG, Y., LI, Q., FENG, Y., LIU, T., & GUO, W. (2018). EPIGENETIC REGULATION OF CANCER PROGRESSION BY EZH2: FROM BIOLOGICAL INSIGHTS TO THERAPEUTIC POTENTIAL. *BIOMARKER RESEARCH*, 6, 10. [HTTPS://DOI.ORG/10.1186/S40364-018-0122-2](https://doi.org/10.1186/S40364-018-0122-2)
- YUAN, B., ZHANG, R., HU, J., LIU, Z., YANG, C., ZHANG, T., & ZHANG, C. (2018). WDR1 PROMOTES CELL GROWTH AND MIGRATION AND CONTRIBUTES TO MALIGNANT PHENOTYPES OF NON-SMALL CELL LUNG CANCER THROUGH ADF/COFLIN-MEDIATED ACTIN DYNAMICS. *INTERNATIONAL JOURNAL OF BIOLOGICAL SCIENCES*, 14(9), 1067-1080. DOI:10.7150/IJBS.23845
- CARÉN, H., FRANSSON, S., EJESKÄR, K., KOGNER, P., & MARTINSSON, T. (2007). GENETIC AND EPIGENETIC CHANGES IN THE COMMON 1P36 DELETION IN NEUROBLASTOMA TUMOURS. *BRITISH JOURNAL OF CANCER*, 97(10), 1416-1424. DOI:10.1038/SJ.BJC.6604032
- NESS, J. K., SKILES, A. A., YAP, E., FAJARDO, E. J., FISER, A., & TAPIPOS, N. (2016). NUC-ERBB3 REGULATES H3K27ME3 LEVELS AND HMT ACTIVITY TO ESTABLISH EPIGENETIC REPRESSION DURING PERIPHERAL MYELINATION. *GLIA*, 64(6), 977-992. DOI:10.1002/GLIA.22977
- TABOLACCI, E., MOSCATO, U., ZALFA, F., BAGNI, C., CHIURAZZI, P., & NERI, G. (2008). EPIGENETIC ANALYSIS REVEALS A EUCHROMATIC CONFIGURATION IN THE FMR1 UNMETHYLATED FULL MUTATIONS. *EUROPEAN JOURNAL OF HUMAN GENETICS*, 16(12), 1487-1498. DOI:10.1038/EJHG.2008.130
- LAMBA, J. K., CAO, X., RAIMONDI, S. C., RAFIEE, R., DOWNING, J. R., SHI, L., ... POUNDS, S. B. (2018). INTEGRATED EPIGENETIC AND GENETIC ANALYSIS IDENTIFIES MARKERS OF PROGNOSTIC SIGNIFICANCE IN PEDIATRIC ACUTE MYELOID LEUKEMIA. *ONCOTARGET*, 9(42), 26711-26723. DOI:10.18632/ONCOTARGET.25475
- SHARMA, A., ALBAHRANI, M., ZHANG, W., KUFEL, C. N., JAMES, S. R., ODUNSI, K., ... KARPF, A. R. (2019). EPIGENETIC ACTIVATION OF POTE GENES IN OVARIAN CANCER. *EPIGENETICS*, 14(2), 185-197. DOI:10.1080/15592294.2019.1581590

REFERENCES

GLOBAL CANCER OBSERVATORY. [HTTPS://GCO.IARC.FR/](https://gco.iarc.fr/)

LHEUREUX S, GOURLEY C, ET AL. (2019). EPITHELIAL OVARIAN CANCER. LANCET. 393: 1240-53.

MOMENIMOVAHED Z, TIZNOBAIK A, ET AL. (2019). OVARIAN CANCER IN THE WORLD: EPIDEMIOLOGY AND RISK FACTORS. INT J WOMENS HEALTH. 11: 287-299.

OZOLS RF, BOOKMAN MA. (2004). FOCUS ON EPITHELIAL OVARIAN CANCER. CANCER CELL. 5:19-24.

CLIBYW, RITLAND S, ET AL. (1993). HUMAN EPITHELIAL OVARIAN CANCER ALLELOTYPE. CANCER RESEARCH. 53. 2393-2398.

SHARMA A, ALBAHRANI M ET AL. (2019). EPIGENETIC ACTIVATION OF POTE GENES IN OVARIAN CANCER. EPIGENETICS. 1559-2308.

BERA TK, FLUER AS, ET AL. (2006). POTE PARALOGS ARE INDUCED AND DIFFERENTIALLY EXPRESSED IN MANY CANCERS. CANCER RESEARCH.66:52-56.

BERA TK, ZIMONJIC DB, ET AL. (2003). POTE, A HIGHLY HOMOLOGOUS GENE FAMILY LOCATED ON NUMEROUS CHROMOSOMES AND EXPRESSED IN PROSTATE, OVARY, TESTIS, PLACENTA, AND PROSTATE CANCER. PNAS. 100(3):16975-16980.

ZHANG G, LIU C, ET AL. (2019). COMBINATORIAL THERAPY OF IMMUNE CHECKPOINT AND CANCER PATHWAYS PROVIDES A NOVEL PERSPECTIVE ON OVARIAN CANCER TREATMENT (REVIEW). ONCOLOGY LETTERS. 17: 2583-2591.

JONES PA, OHTANI H, ET AL. (2019). EPIGENETIC THERAPY IN IMMUNE-ONCOLOGY. NATURE REVIEWS. CANCER.

ZHAO L, SHOU H, ET AL. (2019). EFFECTS OF GINSENOSIDE RG3 ON EPIGENETIC MODIFICATION IN OVARIAN CANCER CELLS. ONCOLOGY REPORTS.41: 3209-3218.

Thank
you